

NOTE

Some Fungi and Water Molds in Waters of Lake Michigan with Emphasis on Those Associated with the Benthic Amphipod *Diporeia* spp.

Bozena Kiziewicz¹ and Thomas F. Nalepa^{2,*}

¹Department of General Biology
Medical University of Bialystok
15-089 Bialystok
Jana Kilinskiego 1, Poland

²Great Lakes Environmental Research Laboratory, NOAA
2205 Commonwealth Blvd.
Ann Arbor, Michigan 48105

ABSTRACT. To determine types of fungi in the water and associated with the benthic amphipod *Diporeia* spp., samples were collected at various depths in Lake Michigan in an area where the *Diporeia* population was in a severe state of decline. No fungi were found associated with living, freshly-dead, or dried *Diporeia* cultured separately from Lake Michigan water. When dead *Diporeia* and other organic substrates (snake skin and hemp seeds) were used to grow fungi in Lake Michigan water, a rich and diverse fungal and water mold community was revealed. A total of 31 species were found, with the most common genera being *Achlya*, *Aphanomyces*, *Myzocytium*, and *Pythium*. In general, species were homogeneously distributed in the water; that is, few differences were found in species richness between nearshore (10–15 m) and offshore (60–80 m) waters, and between near-surface (1 m) and near-bottom waters (1 m off bottom). Sampling occurred during the unstratified period (April and October) to maximize the number of species collected, which may have contributed to the uniform spatial pattern observed. While conclusions must be placed in context with our methods of detection, we found no evidence that a fungal infestation was associated with *Diporeia* in this region of the lake.

INDEX WORDS: Great Lakes fungi, pathogen, disease, macroinvertebrates.

INTRODUCTION

Fungi and fungi-like organisms play a vital role in the decomposition of most organic material in aquatic systems. In particular, specific groups actively mineralize insoluble polysaccharides such as chitin, which comprises the exoskeleton and carapaces of aquatic crustaceans (Batko 1975, Söderhäll *et al.* 1991, Czczuga and Godlewska 2001, Kiziewicz and Kurzatowska 2004). While most fungal groups are saprobionts, other groups are parasitic on the life stages of both invertebrates and vertebrates, including embryos and eggs (Dick 2001a, b). Most of these pathogenetic fungi occur within the division Oomycota (water molds). The

single class Oomycetes within this division have a complex life cycle that includes flagella-bearing zoospores that infect host organisms. While most water molds live freely in the water, they can become associated with host organisms as mostly parasites and predators when environmental conditions change, or when the host is subjected to various stressors (Barron 2003). Various water molds such as *Aphanomyces invadens* can cause massive mortality in fish populations (Noga and Dykstra 1986, Kiryu *et al.* 2002, 2003; Sosa *et al.* 2007), and fungal diseases are widespread in various phytoplankton and crustaceans (Ramaiah 2006).

In this paper we identify some fungal and water molds found in Lake Michigan with an emphasis on species associated with the benthic amphipod *Diporeia* spp. *Diporeia* populations are currently de-

*Corresponding author. E-mail: Thomas.Nalepa@noaa.gov

clining in all the Great Lakes except Lake Superior, and large areas are now completely devoid of this organism (Nalepa *et al.* 2006a). While the disappearance of *Diporeia* has been linked to the introduction and spread of dreissenid mussels (*Dreissena polymorpha* and *Dreissena bugensis*), exact causes for the decline have been difficult to determine (Nalepa *et al.* 2005, Nalepa *et al.* 2006b). By examining fungi associated with *Diporeia* in an area where populations are declining, we explore the possibility that a water-borne fungus may be implicated in observed declines. Pathogens are common in amphipods and are known to reduce or limit populations (Johnson 1985, 1986), or to play a major role in structuring communities (MacNeil *et al.* 2003). A dramatic decline in the amphipod *Corophium violator* in the Baltic Sea was attributed to a yeast pathogen (Seegerstrale 1960). In the Great Lakes, Messick *et al.* (2004) used histological techniques to determine the prevalence of parasites and pathogens associated with *Diporeia* from Lakes Michigan and Huron. Incidence levels of yeast-like organisms and fungi were low, only 0.4%, and levels in areas of declining populations were not different from those in areas where populations were stable. To supplement this work, we further examine fungi and water molds associated with *Diporeia* by using the “bait” method. In this method, organic matter is used as substrate for the cultured growth of fungi; hence, the method may detect the presence of fungal taxa not easily observed using histological techniques. In addition to determining the presence of fungi associated with *Diporeia*, we document those taxa present in nearshore and offshore waters of Lake Michigan that readily become associated with organic material.

MATERIAL AND METHODS

Sampling sites were located in Lake Michigan at various depths off Muskegon, MI (Fig. 1). *Diporeia* were collected with a Ponar grab at 60 m (43° 12.00' N, 86° 27.37' W), and at 80 m (43° 12.00' N, 86° 29.57' W) in April and October, 2004, respectively. Sampling was conducted in the spring and fall since aquatic fungi grow best at colder temperatures (Srivastava 1967, Khulbe and Bhargava 1977). The *Diporeia* population at the 60-m site was in a severe state of decline at the time of sampling in April, and individuals were no longer found at this site by October. Hence, to collect *Diporeia* in October, we sampled progressively offshore until individuals were found, and that was not

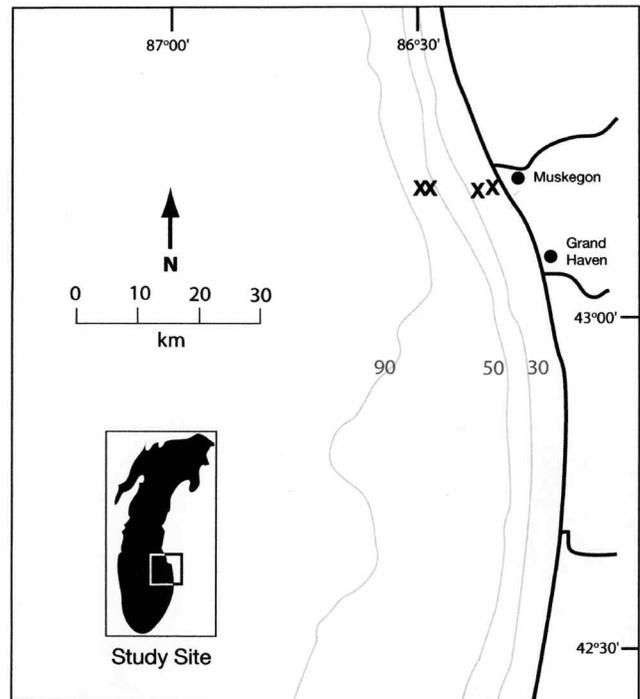


FIG. 1. Location of sampling sites (X) in south-eastern Lake Michigan off Muskegon, MI. Sites were located at 10–15 m, 25 m, 60 m, and 80 m. Depth contours are 30, 50, and 90 m.

until we reached the 80 m depth. Intact Ponar grab samples (organisms, sediment, and overlying water) were placed into plastic bags, then placed into a cooler, iced, and brought back to the laboratory and kept at 4°C. Water samples were also collected at each site with a rinsed, 1-L Niskin bottle from three vertical depths: 1 m, 4 m, and 1 m off the bottom. In addition to collecting water samples at sites where we collected *Diporeia*, we also collected water at two additional sites in both April and October—one site was located at 10–15 m (43° 11.49' N, 86° 20.64' W) and the other site was located at 25 m (43° 12.00' N, 86° 22.67' W). *Diporeia* had already disappeared from these two shallower sites several years earlier (Nalepa unpublished data).

Within 24 h of collection, 10–20 *Diporeia* of various sizes (~ 4–7 mm) were removed from the sediment samples, dried in a dessicator at room temperature for 48 h, and then placed in clean, sealed vials. These dried specimens were sent via overnight express to Medical University, Bialystok, Poland for analysis of attached fungi. Also sent were the water samples collected from each site and vertical depth, as well as 10–20 live specimens

from each collection location. The latter individuals were shipped in rinsed, insulated thermos bottles filled with Lake Michigan water collected from the hypolimnion.

Fungi were isolated using the bait method in which organic material is used as substrate for fungal culture (Seymour and Fuller 1987). In addition to *Diporeia*, hemp seeds (*Cannabis sativum*) and snake skin (*Natrix natrix*) were also used as substrate for fungal growth. Both hemp seeds and snake skin are standard substrates used in culturing fungi for identification. Each of the three bait items was placed separately into 1-L containers with Lake Michigan water. Living and freshly-dead *Diporeia* from the thermos bottles, and previously dried *Diporeia*, were placed into petri dishes containing distilled water. All containers were covered with glass plates to partly protect the water from outside bacteria and then incubated for approximately 1 month at room temperatures. During this period, microscopically-determined mycelia were removed from each of the substrates and transferred to sterilized petri dishes containing distilled water. Several preparations were made for each sample, and aquatic fungi were identified using the vegetative organs (shape and size of the hyphae), the asexual organs (shape of sporangium and spores), and the generative organs (structure of the oogonium, oosporangium, and antheridium). Observations and measurements were made on colonies developed on these baits. Incubation at room temperature proved to be adequate for vigorous growth of every representative we isolated. Keys used for fungi identification are given in Sparrow (1960), Bedenek (1972), Batko (1975), Fassatiova (1983), Seymour and Fuller (1987), Dick (1969, 1990), and Pystina (1998).

Fungi found in Lake Michigan water exclusive of the bait method were determined using solid media culture techniques (agar-medium method). Water (1 mL) from each of the sites and vertical depths was sowed onto culture plates containing Sabouraud glucose agar with Gentamicin and chloramphenicol 2 (SGCs; BioMerieux, France) and then incubated for 3–5 days at 28°C. Observed fungi were further transferred to plates with Sabouraud agar and incubated at room temperature for 5 days, during which developing colonies were counted, examined, and identified (Zaremba and Borowski 2001). In this way, pure fungi developed with the structures needed for identification. Species identification was based on morphological and biometrical characters of the mycelium and conidia (Fassatiova 1983, Zaremba and Borowski 2001).

For purposes of analysis, the sites were divided into three depth categories: 10–15 m, 25 m, and 60–80 m. Independence in the number of species collected between the 2 months (April, October), three sites, and three vertical depths (1 m, 4 m, 1 m off bottom) were tested using a G-test (log-likelihood) within a $2 \times 3 \times 3$ contingency table (Sokal and Rohlf 1969).

RESULTS

No fungi (mycelia) were found on living, freshly-dead, or dried *Diporeia* placed in distilled water indicating that, at the time of collection, no fungi able to grow under the described circumstances were associated with these individuals. When using organic bait (snake skin, hemp seeds, and dried *Diporeia*) and the agar-medium method to culture fungi from Lake Michigan water, we identified 18 genera and 31 fungal species (Table 1). Of these, 18 were found on snake skin, 12 on *Diporeia*, 5 on hemp seeds, and 3 on Sabouraud glucose agar. The most common genera were fungi within the class Oomycetes: *Achlya* with seven species found, *Pythium* with four species, and *Aphanomyces* and *Myzocyttium* and with three species each. Of these major genera, some preferences were noted in the substrate selected. For instance, the four *Pythium* and three *Aphanomyces* species grew exclusively on snake skin, and all seven of the *Achlya* species grew on *Diporeia*, with three of these growing on *Diporeia* exclusively.

Although sampling date, site, and water-column depth jointly lacked independence ($0.05 > P > 0.01$), consistent patterns in the distribution of total species were not apparent across these factors (Table 1). The lack of independence was likely a result of the greater number of species (12) found at 1-m water depth at the 60-m site in April as compared to the other date \times site \times water depth combinations (maximum of six species). Five of the seven species of *Achlya* were only found on that date and location. The total number of species was generally similar at the three sites (15, 11, and 15 species at 10–15 m, 25 m, and 60–80 m sites), and vertically within the water column (16, 14, and 11 species at 1m, 4 m, and 1 m off the bottom).

DISCUSSION

A number of studies have examined the occurrence of fungal communities in Great Lakes waters (Paterson 1967, Qureshi and Dutka 1974, El-

Shaarawi *et al.* 1977, Sherry 1986, Kwasniewska 1988), but it is difficult to compare our results to those of these previous studies because of different techniques of collection, culturing mediums, and levels of identification. Kwasniewska (1988) used a select agar medium and identified seven dominant genera of yeasts and filamentous fungi in Lake St. Clair, of which only one was found in our Lake Michigan samples (*Fusarium*). Likewise, Qureshi and Dutka (1974) used various agar mediums and found 25 genera in Lake Ontario waters off the Niagara River mouth, of which only five were found in Lake Michigan (*Alernaria*, *Aspergillus*, *Fusarium*, *Geotrichum*, and *Penicillium*). We identified 18 genera when using both the bait and the agar-medium methods, but only three were identified using the latter method. Hence, when just considering the agar-culture method, it appears that fewer genera were found in the Muskegon area of Lake Michigan relative to these previous studies in Lake St. Clair and the Niagara River area of Lake Ontario. It should be noted, however, that these earlier studies were mostly conducted in the 1970s when, because of greater nutrient loads and absence of *Dreissena*, more organic material was likely present in the water column. Fungi tend to be more abundant and diverse in areas subjected to organic enrichment (Quershi and Dutka 1974).

Based on occurrences in other water bodies that have used the "bait" method, mostly lakes in Europe, 18 of the 31 species found in Lake Michigan might be considered rare, while the other 13 might be considered common (see Table 1; B. Kiziewicz, unpublished data). The genera with the most species in Lake Michigan, all within the class Oomycetes, were *Achlya*, *Aphanomyces*, *Myzocytium*, and *Pythium*. All seven species of *Achlya* grew on dead *Diporeia* when it was used as a substrate. These species typically colonize chitin-and-keratin containing substrates such as insect exuviae, and dead fish and amphipods (Czeczuga and Godlewska 1994, Czeczuga *et al.* 2004). Studies have also shown that some of these species (*A. diffusa*, *A. polyandra*, *A. prolifera*, and *A. proliferoides*) can be parasitic on fish eggs and adults (Srivastava and Srivastava 1977, Czeczuga *et al.* 2002). The species of *Aphanomyces* and *Pythium* found in Lake Michigan are mostly saprobiotic, living on decayed animal and plant remains. The exception is *Aphanomyces laevis*, which is a widely-occurring species that is parasitic on fish (Dykstra *et al.* 1986). *Myzocytium zoophthorum* was the most frequently encountered species; this

species along with other species of this genus are described as parasitic on small invertebrates, mostly rotifers (Batko 1975). Another species found in Lake Michigan, *Fusarium moniliformae*, has been described as an opportunistic pathogen of crustaceans (Ramaiah 2006).

The main purpose of this study was to determine fungi and water molds associated with *Diporeia*. Population densities of *Diporeia* in the Lake Michigan study area off Muskegon have been in a steady state of decline since the late 1990s. The decline of *Diporeia* occurred first in shallow, nearshore waters and then proceeded offshore, which is consistent with the offshore expansion of dreissenid mussels (Nalepa *et al.* 2006a). The population density of *Diporeia* at a 45-m site off Muskegon was 10,000/m² in early 1997, but declined to near 0/m² by the end of 2002 (Nalepa *et al.* 2006b). At our 60-m site, *Diporeia* were present in spring 2004, but had disappeared by fall 2004. Densities at our 80-m site have not been regularly monitored, but the population was likely in the state of decline at the time samples were taken. For example, densities at a nearby 100-m site were 4,400/m² in 2002, 1,040/m² in 2004, and 4/m² in 2005 (Nalepa unpublished data). Given these population declines, it is noteworthy therefore that no fungi were found associated with living, freshly-dead, or dried *Diporeia* when cultured apart from Lake Michigan water. Instead, all the fungal species found on *Diporeia* developed after individuals were cultured in Lake Michigan water, indicating fungi were present in the water, but not associated with *Diporeia*.

While relative abundances of fungi were not determined, some insights into distributions can be derived from the relative number of species found. Unexpectedly, gradients related to sampling depth (distance offshore) and vertically in the water-column were not apparent. That is, more species were expected at shallow sites compared to deeper ones, and more near bottom than near surface in the water column. Aquatic fungi utilize a variety of organic substrates for growth, and likely enter lakes through watershed inputs (Dick 1971). Thus, more fungal species might be expected in nearshore areas and near the bottom sediments where organic matter content would be highest. Since, as noted, aquatic fungi grow best at colder temperatures (Srivastava 1967, Khulbe and Bhargava 1977), we collected during the unstratified period in April and October to maximize the number of species found. Sampling during this period of uniform mixing may have contributed to the lack of observed gradients both hori-

zontally and vertically within the water column. The exception to this pattern of homogenous distributions was the relatively high number of *Achlya* species found near the surface at the 60-m site in April. Species of this genus are chitinophilic and readily colonize dead amphipods. The decreasing *Diporeia* population at this site may have contributed to the occurrence of these species, but *Achlya* were not found throughout the water column and were not found in October at the 80-m site where the *Diporeia* population was also likely decreasing.

In summary, this study seems to confirm the finding of Messick *et al.* (2004) that fungi associated with living or freshly-dead *Diporeia* in Lake Michigan were absent or rare. While a rich and diverse community of fungi and water mold species was found in Lake Michigan waters when using various substrates as organic "bait" (including freshly-dead *Diporeia*), no fungi could be cultured using *Diporeia* as a substrate separate from Lake Michigan water. Within the context of our detection methods, therefore, there was no indication that a fungal infection was primarily the cause of the population decline. While some results suggest that a pathogen/disease may be a potential reason for the decline of *Diporeia* in the Great Lakes (Dermott *et al.* 2005), further, more-detailed studies are needed to explore this possibility.

ACKNOWLEDGMENTS

The authors thank the crew of the R/V *Laurentian* for their assistance and cooperation during field operations and D.L. Fanslow for collection of specimens. We also thank Prof. Dr. Ludwik Zmudzinski (deceased) of Pomeranian Academy of Education, Slupsk, Poland for thoughtful discussions and help in initiating this study. This is GLERL Contribution No. 1477.

REFERENCES

- Barron, G.L. 2003. Predatory fungi, wood decay, and the carbon cycle. *Biodiversity* 4:3–9.
- Batko, A. 1975. Zarys hydromikologii [Hydromycology—an overview]. PWN, Warszawa.—[in Polish]
- Bedenek, T. 1972. Fragmenta Mycologica. I. Some historical remarks of the development of "hairbaiting" of Toma-Karling-Vanbreuseghem (the To-Kava hairbaiting method). *Mycopathol. Appl.* 68:104–106.
- Czczuga, B., and Godlewska, A. 1994. Aquatic fungi growing on substrates containing chitin. *Acta Mycol.* 29:189–200.
- _____, and Godlewska, A. 2001. Aquatic insects as vectors of aquatic zoosporic fungi parasitic on fishes. *Acta Ichtyol. Piscat.* 31:87–104.
- _____, Kozłowska, M., and Godlewska, A. 2002. Zoosporic aquatic fungi growing on dead specimens of 29 freshwater crustacean species. *Limnologica* 32:180–193.
- _____, Kiziewicz, B., and Gruszka, P. 2004. *Pallasea quadrispinosa* GO Sars specimens as vectors of aquatic zoosporic fungi parasiting on fish. *Pol. J. Environ. Stud.* 13:361–366.
- Dermott, R., Munawar, M., Bonnell, R., Carou, S., Niblock, H., Nalepa, T.F., and Messick, G.A. 2005. Preliminary investigations into causes of the disappearance of *Diporeia* from Lake Ontario. In *Proceedings of a workshop on the dynamics of lake whitefish (Coregonus clupeaformis) and the amphipod Diporeia spp. in the Great Lakes*. L. C. Mohr and T. F. Nalepa, eds., pp. 203–232. Great Lakes Fish Comm. Tech. Rep 69.
- Dick, M.W. 1969. Morphology and taxonomy of the Oomycetes, with special reference to Saprolegniaceae, Leptomitaceae and Pythiaceae. I. Sexual reproduction. *New Phytologist* 68:751–775.
- _____. 1971. Ecology of Saprolegniaceae in lentic and littoral muds with a general theory of fungi in the lake ecosystem. *J. Gen. Microbiol.* 65:325–327.
- _____. 1990. *Keys to Pythium*. Coll. Estate Manag. Whiteknights, Reading, U.K.
- _____. 2001a. The Peronosporomycetes. In *The Mycota VII. Part A. Systematics and evolution*. D.J. McLaughlin, E.G. McLaughlin, P.A. Lemke, eds., pp. 39–72. Berlin: Springer-Verlag.
- _____. 2001b. *Straminipilous Fungi*. Kluwer Academic Publishers.
- Dykstra, M.J., Noga, E.J., Levine, J.F., Moye, D.W., and Hawkins, J.H. 1986. Characterization of the *Aphanomyces* species involved with ulcerative mycosis (UM) in menhaden. *Mycologia* 78:664–672.
- El-Shaarawi, A., Qureshi, A.A., and Dutka, B.J. 1977. Study of microbiological and physical parameters in Lake Ontario adjacent to the Niagara River. *J. Great Lakes Res.* 3:196–203.
- Fassatiowa, O. 1983. Grzyby mikroskopowe w mikrobiologii technicznej [Microscopic fungi in technical microbiology]. WN-T., Warszawa.—[in Polish]
- Johnson, P.T. 1985. Parasites of benthic amphipods: microsporidians of *Ampelisca agassizi* (Judd) and some other gammarideans. *U.S. Fish. Wildl. Serv. Fish. Bull.* 83:497–505.
- _____. 1986. Parasites of benthic amphipods: dinoflagellates (Duboscquodinida: Syndinidae). *U.S. Fish. Wildl. Serv. Fish. Bull.* 84:605–614.
- Khulbe, R.D., and Bhargava, K.S. 1977. Distribution and seasonal periodicity of water molds in some lakes in Naini Tal hills, India. *Hydrobiologia* 54:67–72.
- Kiryu, Y., Shields, J.D., Vogelbein, W.K., Zwerner,

- D.E., Kator, H., and Blazer, V.S. 2002. Induction of skin ulcers in Atlantic menhaden by injection and aqueous exposure to the zoospores of *Aphanomyces invadans*. *J. Aquat. Anim. Health* 14:11–24.
- Kiryu, Y., Shields, J.D., Vogelbein, W.K., Zwerner, D.E., Kator, K., and Blazer, V.S. 2003. Infectivity and pathogenicity of the oomycete *Aphanomyces invadans* in Atlantic menhaden *Brevoortia tyrannus*. *Dis. Aquat. Org.* 54:135–146.
- Kiziewicz, B., and Kurzatowska, A. 2004. Aquatic fungi and fungus-like organisms isolated from surface waters situated near Białystok in Podlasie Province of Poland using the insect *Notonecta glauca* L. as bait. *Mycol. Balcan.* 1:117–123.
- Kwasniewska, K. 1988. Horizontal distribution and density of yeasts and filamentous fungi in Lake St. Clair water. *J. Great Lakes Res.* 14:438–443.
- MacNeil, C., Dick, J.T.A., Hatcher, M.J., Terry, R.S., Smith, J.E., and Dunn, A.M. 2003. Parasite-mediated predation between native and invasive amphipods. *Proc. Royal. Soc. Lond. B.* 270:1309–1314.
- Messick, G.A., Overstreet, R.M., Nalepa, T.F., and Tyler, S. 2004. Prevalence of parasites in amphipods *Diporeia* spp. from Lakes Michigan and Huron, USA. *Dis. Aquat. Org.* 59:159–170.
- Nalepa, T.F., Fanslow, D.L., and Messick, G. 2005. Characteristics and potential causes of declining *Diporeia* spp. populations in southern Lake Michigan and Saginaw Bay, Lake Huron. In *Proceedings of a workshop on the dynamics of lake whitefish (Coregonus clupeaformis) and the amphipod Diporeia spp. in the Great Lakes*. L.C. Mohr and T.F. Nalepa, eds., pp. 157–188. Great Lakes Fish. Comm. Tech. Rep. 66.
- , Rockwell, D.C., and Schloesser, D.W. 2006a. *Disappearance of the amphipod Diporeia spp. in the Great Lakes: Workshop summary, discussion, and recommendations*. NOAA Technical Memorandum GLERL-136. NOAA, Great Lakes Environmental Research Laboratory, Ann Arbor, MI.
- , Fanslow, D.L., Foley, A.J., III, Lang, G.A., Eadie, B.J., and Quigley, M.A. 2006b. Continued disappearance of the benthic amphipod *Diporeia* spp. in Lake Michigan: is there evidence for food limitation? *Can. J. Fish. Aquat. Sci.* 63:872–890.
- Noga, E.J., and Dykstra, M.J. 1986. Oomycete fungi associated with ulcerative mycosis in menhaden, *Brevoortia tyrannus* (Latrobe). *J. Fish. Dis.* 9:47–53.
- Paterson, R.A. 1967. Benthic and planktonic phycomycetes from Northern Michigan. *Mycologia* 59:405–416.
- Pystina, K.A. 1998. Genus *Pythium* Pringhs. Nauka, St. Petersburg, Russia.
- Qureshi, A.A., and Dutka, B.J. 1974. A preliminary study on the occurrence and distribution of geo-fungi in Lake Ontario and the Niagara River. In *Proc. 17th Conf. Great Lakes Res.*, pp. 653–662. Internat. Assoc. Great Lakes Res.
- Ramaiah, N. 2006. A review on fungal diseases of algae, marine fishes, shrimps and corals. *Ind. J. Mar. Sci.* 35:380–387.
- Seegerstrale, S.G. 1960. Fluctuations in the abundance of benthic animals in the Baltic area. *Commentat. Biol. Soc. Sci. Fenn.* 23:1–19.
- Seymour, R.L., and Fuller, M.S. 1987. Collection and isolation of water molds (Saprolegniaceae) from water and soil. In *Zoosporic Fungi in Teaching and Research*. M.S. Fuller and A. Jaworski, eds., pp. 125–127. Athens: Southern Publishing.
- Sherry, J.P. 1986. Temporal distribution of geoaquatic fungi at a nearshore station in Lake Ontario. *J. Great Lakes Res.* 12:221–224.
- Söderhäll, K., Dick, M.W., Clark, G., Furst, M., and Constantinescu, O. 1991. Isolation of *Saprolegnia parasitica* from the crayfish *Astacus leptodactylus*. *Aquaculture* 92:121–125.
- Sokal, R.R., and Rohlf, F.J. 1969. *Biometry*. W. H. Freeman and Co., San Francisco.
- Sosa, E.R., Landsberg, J.H., Stephenson, C.M., Forstchen, A.B., Vandersea, M.W., and Litaker, R.W. 2007. *Aphanomyces invadans* and ulcerative mycosis in estuarine and freshwater fish in Florida. *J. Aquat. Anim. Health* 19:14–26.
- Sparrow, F.K. 1960. *Aquatic Phycomycetes 2nd ed.*, University of Michigan Press, Ann Arbor, MI.
- Srivastava, G.C. 1967. Ecological studies on some aquatic fungi of Gorakhpur, India. *Hydrobiologia*. 30:385–404.
- , and Srivastava, R.C. 1977. Host range of *Achlya proliferata* (Nees) de Bary on certain freshwater teleosts. *Mycopathologia* 61:61–62.
- Zaremba, L., and Borowski, J. 2001. Mikrobiologia lekarska [Medical microbiology]. PZWŁ, Warszawa. -[in Polish]

Submitted: 5 November 2007

Accepted: 2 July 2008

Editorial handling: David R. Barton